




# REVISTA ARGENTINA DE MICROBIOLOGÍA

[www.elsevier.es/ram](http://www.elsevier.es/ram)



## ORIGINAL ARTICLE

# Antimicrobial susceptibility of *Clostridioides difficile*. An Argentinian multicenter study of isolates from human patients

Raquel Rollet <sup>a,b,i,\*</sup>, Daniela Vaustat<sup>a,b,i</sup>, Mirta Litterio<sup>c,i</sup>, Liliana Castello<sup>d,i</sup>,  
Liliana Fernández-Canigia<sup>e,i</sup>, Claudia Barberis<sup>f,i</sup>, María Cristina Legaria<sup>f,i</sup>,  
Natalia Azula<sup>g,i</sup>, María Laura Maldonado<sup>c,i</sup>, Gerardo Laube<sup>b</sup>, Silvia Carla Predari<sup>d,i</sup>,  
María Adelaida Rossetti<sup>h,i</sup>

<sup>a</sup> Hospital de Enfermedades Infecciosas Dr. Francisco Javier Muñiz, Unidad de Bacteriología, Ministerio de Salud, Ciudad Autónoma de Buenos Aires, Argentina

<sup>b</sup> Instituto Universitario de Ciencias de la Salud, Fundación HA Barceló, Facultad de Medicina, Cátedra de Microbiología, Ciudad Autónoma de Buenos Aires, Argentina

<sup>c</sup> Hospital de Pediatría Prof. Dr. Juan P. Garrahan, Servicio de Microbiología, Ciudad Autónoma de Buenos Aires, Argentina

<sup>d</sup> Universidad de Buenos Aires, Instituto de Investigaciones Médicas Alfredo Lanari, Departamento de Microbiología, Ciudad Autónoma de Buenos Aires, Argentina

<sup>e</sup> Hospital Alemán, Sector Microbiología, Laboratorio Central, Ciudad Autónoma de Buenos Aires, Argentina

<sup>f</sup> Universidad de Buenos Aires, Facultad de Farmacia y Bioquímica, Departamento de Bioquímica Clínica, Cátedra de Microbiología Clínica, Ciudad Autónoma de Buenos Aires, Argentina

<sup>g</sup> Centro de Educación Médica e Investigaciones Clínicas Norberto Quirno (CEMIC), Laboratorio de Microbiología, Ciudad Autónoma de Buenos Aires, Argentina

<sup>h</sup> Hospital Interzonal General de Agudos Presidente Perón, Unidad de Microbiología, Avellaneda, Provincia de Buenos Aires, Argentina

<sup>i</sup> Subcomisión de Bacterias Anaerobias, Sociedad Argentina de Bacteriología, Micología y Parasitología Clínica, Asociación Argentina de Microbiología, Argentina

Received 26 December 2024; accepted 17 September 2025

### KEYWORDS

*Clostridioides  
difficile*;  
Antimicrobial  
susceptibility;  
Multicenter;  
Multidrug resistance;  
Argentina

**Abstract** *Clostridioides difficile* is an etiological agent of diarrhea, and the use of antibiotics is one of the main risk factors for infection. Antimicrobials used for treatment are vancomycin (VAN), metronidazole (MET), and fidaxomicin. Resistant strains have been detected, exhibiting regional and institutional differences. The aim of this work was to determine the susceptibility profile of *C. difficile* clinical isolates to 14 antimicrobials, and to compare resistance among participating centers. A total of 208 consecutive isolates recovered from seven Argentinian hospitals between January 2018 and March 2020 were studied. MIC was determined by the agar dilution method (CLSI-M100 29ED). Azithromycin (AZM), clindamycin (CLI), ertapenem (ETP),

\* Corresponding author.

E-mail address: [rarollet@yahoo.com](mailto:rarollet@yahoo.com) (R. Rollet).

<https://doi.org/10.1016/j.ram.2025.09.006>

0325-7541/© 2025 The Authors. Published by Elsevier España, S.L.U. on behalf of Asociación Argentina de Microbiología. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Please cite this article as: R. Rollet, D. Vaustat, M. Litterio et al., Antimicrobial susceptibility of *Clostridioides difficile*. An Argentinian multicenter study of isolates from human patients, Revista Argentina de Microbiología, <https://doi.org/10.1016/j.ram.2025.09.006>

imipenem (IMI), levofloxacin (LEV), linezolid (LNZ), meropenem (MER), metronidazole (MET), moxifloxacin (MOX), piperacillin-tazobactam (PTZ), rifaximin (RFX), teicoplanin (TEI), tigecycline (TGC), and VAN, were tested. The results were analyzed with SPSS 21.0. Chi-square was used to compare data, and statistical significance was set at  $p < 0.05$ . Susceptibility percentages were as follows: VAN, TEI, and MET, 100%; TGC, 97.6%; PTZ, 96.2%; LNZ, 95.2%; MER, 99.5%; ETP, 60.9%; IMI, 42.8%; RFX, 55.6%; LEV, 48.6%; MOX, 46.1%; CLI, 29.9%; and azithromycin, 17.8%. Significant differences in resistance among centers were observed for: RFX (16.7%–91.7%), CLI (41.2%–86.1%), MOX (22.9%–97.2%), IMI (0%–55.6%), and azithromycin (62.5%–97.2%). Multidrug resistance (MDR) was detected in 80 isolates (38.5%), of which 63 (78.7%) were resistant to three families of antimicrobial agents and 17 (21.3%) were resistant to four. The most frequent combinations were RFX–MOX–CLI, present in 48 (60.0%) isolates, and RFX–IMI–MOX–CLI in 17 (21.3%) isolates. VAN, TEI, and MET were the most active antimicrobials *in vitro* against *C. difficile* strains. MER was the most active carbapenem, whereas IMI was the least active. We highlight the differences across institutions that could reflect epidemiological characteristics, and/or the dissemination of clones in each institution.

© 2025 The Authors. Published by Elsevier España, S.L.U. on behalf of Asociación Argentina de Microbiología. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## PALABRAS CLAVE

*Clostridioides difficile*;  
Sensibilidad  
antibiótica;  
Multicéntrico;  
Multirresistencia;  
Argentina

## Sensibilidad antibiótica de *Clostridioides difficile*. Un estudio argentino multicéntrico de aislados de pacientes humanos

**Resumen** *Clostridioides difficile* causa diarreas, y el uso de antibióticos es uno de los principales factores de riesgo. Los tratamientos se realizan con vancomicina (VAN), metronidazol (MET) y fidaxomicina. Se detectaron cepas resistentes con diferencias regionales e institucionales. Los objetivos de este trabajo fueron determinar la sensibilidad de aislados clínicos de *C. difficile* frente a 14 antimicrobianos y comparar la resistencia entre centros. Se estudiaron 208 aislados consecutivos de 7 hospitales argentinos entre enero de 2018 y marzo de 2020. Se determinó la CIM por dilución en agar (CLSI-M100 29ED) de azitromicina (AZM), clindamicina (CLI), ertapenem (ETP), imipenem (IMI), levofloxacina (LEV), linezolid (LNZ), meropenem (MER), MET, moxifloxacina (MOX), piperacilina-tazobactam (PTZ), rifaximina (RFX), teicoplanina (TEI), tigeciclina (TGC) y VAN. Se utilizó chi-cuadrado para comparar los datos y se asignó significación estadística a  $p < 0,05$  (SPSS21.0). Los porcentajes de sensibilidad fueron: VAN, TEI y MET, 100%; TGC, 97,6%; PTZ, 96,2%; LNZ, 95,2%; MER, 99,5%; ETP, 60,9%; IMI, 42,8%; RFX, 55,6%; LEV, 48,6%; MOX, 46,1%; CLI, 29,9%, y AZM, 17,8%. Entre centros, se encontraron diferencias significativas en la resistencia: RFX (16,7%-91,7%), CLI (41,2%-86,1%), MOX (22,9-97,2%), IMI (0-55,6%) y AZM (62,5-97,2%). Se detectó multirresistencia en 80 (38,5%) aislados. VAN, TEI y MET fueron los antimicrobianos más activos frente a *C. difficile*. MER fue el carbapenem más activo e IMI el menos activo. Destacamos las diferencias entre instituciones, que podrían reflejar características epidemiológicas o de diseminación clonal en cada institución.

© 2025 Los Autores. Publicado por Elsevier España, S.L.U. en nombre de Asociación Argentina de Microbiología. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Introduction

*Clostridioides difficile* is a toxigenic Gram-positive rod, spore-forming anaerobe that causes intestinal diseases. *C. difficile* infection (CDI) is mediated by the production of two toxins: an enterotoxin (toxin A), encoded by the *tcdA* gene, and a cytotoxin (toxin B) encoded by the *tcdB* gene. Some strains can produce a third unrelated binary toxin, which is encoded by the *cdtA-cdtB* genes and has no clearly defined role in humans.

*C. difficile* is a leading cause of diarrhea in hospitalized patients worldwide. It is the etiological agent of 25–30%

of hospital-acquired diarrhea; however, CDI also occurs in community patients.

*C. difficile* causes 15–25% of episodes of diarrhea associated with antibiotic treatment. Exposure to antimicrobials plays a significant role in the pathogenesis of CDI. Antibiotics produce disruption of the gut microbiota and create an environment susceptible to *C. difficile* colonization and subsequent CDI<sup>4</sup>. This event is favored by the high level of antibiotic resistance detected in *C. difficile* clinical isolates<sup>41</sup>.

Antibiotic administration is the most common risk factor for CDI<sup>49</sup>; consequently, almost all antimicrobial agents

have been associated with the development of CDI. However, agents such as amoxicillin-clavulanic acid, CLI, cephalosporins, and fluoroquinolones pose a higher potential risk<sup>2,4,8,45</sup>. Other important risk factors are hospitalization, residence in chronic care institutions, and environmental colonization<sup>4,8,45</sup>.

Clinical manifestations range from mild diarrhea to life-threatening toxic megacolon and pseudomembranous colitis, sepsis and death<sup>45,49</sup>. Additionally, up to 20% of patients with CDI with a favourable outcome experience a recurrent episode within four weeks of completing the treatment. Among them, the risk of a third episode rises to 60%.

The antibiotics used for the treatment of CDI are vancomycin (VAN), metronidazole (MET), and fidaxomicin<sup>25,41,46</sup>; nevertheless, the latter is not available in all countries. Thus, in Argentina, MET and VAN are the most prescribed agents for the treatment of this infection<sup>4</sup>. Teicoplanin (TEI), tigecycline (TGC), and nitazoxanide are alternative drugs, and in the absence of response to treatment or recurrences, linezolid (LNZ) or rifaximin (RXM) could be added<sup>4,5</sup>. However, the emergence of antibiotic resistance and multidrug resistance has been reported; therefore, periodic monitoring is vital for appropriate therapy<sup>13</sup>. Due to regional and institutional variations<sup>49</sup>, knowledge of the resistance profiles of locally circulating *C. difficile* strains is crucial. In Argentina few antibiotic susceptibility studies involving human clinical isolates of *C. difficile* have been conducted<sup>28,30,38</sup>.

The present study aimed to determine antimicrobial susceptibility patterns of 208 *C. difficile* strains isolated from patients with CDI against 14 antibiotics and to compare resistance across the participating centers.

## Materials and methods

### Bacterial isolates

A total of 208 consecutive isolates of *C. difficile* were studied. They were obtained from seven hospital centers from the Metropolitan Area of Buenos Aires, Argentina, six from Buenos Aires City (CE, HA, HC, HG, HM, IL), and one from Buenos Aires Province (HP), and were collected between January 2018 and March 2020. The distribution of isolates across the participating centers was as follows: CE 32, HA 33, HC 31, HG 35, HM 36, HP 36, IL 5.

They were isolated from stool samples of patients with diarrhea and a diagnosis of CDI. The diagnosis was performed according to a two-step algorithm<sup>4</sup>. In all cases, GDH and toxins were detected using a membrane enzyme immunoassay kit (C. Diff Quik Chek Complete™, TECHLAB), and toxigenic cultures were used for the second step when necessary.

Then, in order to obtain the isolates for this study, a toxigenic culture was done on all samples that resulted positive. Following alcohol shock-based selection of spores, isolation of *C. difficile* was performed. Equal parts of feces and 98% ethanol were mixed, vortexed and incubated at room temperature for 20 min. Afterwards, 10 µl of the mix was poured onto blood agar plates, which were incubated in jars under anaerobic conditions at 35 °C for 48–72 h.

Suspicious colonies were identified by their characteristic colony morphology, Gram stain, biochemical tests, and mass spectrometry systems such as MALDI-TOF MS, BD Bruker Biotyper™ (Bruker Daltonics, Bremen, Germany) or Vitek™ MS (bioMérieux, Marcy-L'Etoile, France), when necessary. Toxin production was detected using the C. Diff Quik Chek Complete™ kit, TECHLAB.

The strains were stored at –20 °C in brain heart infusion broth plus 15% glycerol. To recover the isolates, two subcultures were performed onto blood agar plates, and incubated under anaerobic conditions at 35 °C for 48 h.

All the isolates were subsequently sent to the research laboratory at the Facultad de Medicina of the Fundación Barceló, where antibiotic susceptibility testing was performed.

### Antimicrobial susceptibility tests

Minimum inhibitory concentrations (MICs) were determined by agar dilution method according to CLSI-M100 29th edition<sup>9</sup>. Tests were performed on Brucella agar supplemented with haemin (5 µg/ml), vitamin K1 (1 µg/ml), and laked horse blood (5% v/v).

The antibiotics tested were of certified potency, and their respective concentration ranges in µg/ml were: azithromycin (AZM), 0.12–128; clindamycin (CLI), 0.12–64; ertapenem (ETP), 0.12–64; imipenem (IMI), 0.12–64; levofloxacin (LEV), 0.12–128; LNZ, 0.03–16; meropenem (MER), 0.12–64; MTZ, 0.12–128; moxifloxacin (MOX), 0.12–64; piperacillin-tazobactam (PTZ), 0.25–512; RFX, 0.002–128; TEI, 0.008–16; TGC 0.06–4, and VAN, 0.03–16.

MICs were recorded after 48 h of incubation and values were interpreted according to CLSI-M100 33rd edition<sup>10</sup>. For those agents whose breakpoints were not defined by CLSI, EUCAST or published data<sup>15,16,37,49</sup> were adopted.

Breakpoints for MTZ, CLI, IMI, MER, ETP, MOX, and PTZ were those defined by CLSI-M100 33rd edition guidelines for anaerobic bacteria<sup>10</sup>. The breakpoints for VAN and TGC were defined according to EUCAST epidemiological cut-off (ECOFF) values for *C. difficile*<sup>15,16</sup>, for RFX resistant breakpoint was >32 µg/ml<sup>37</sup>, for LEV, >8 µg/ml<sup>49</sup>. The values for TEI, LNZ and AZM were >2 µg/ml, >4 µg/ml and >2 µg/ml, respectively, and were adopted from those defined for *Staphylococcus aureus*<sup>15</sup>.

Quality control strains included *C. difficile* ATCC 700057, *Bacteroides fragilis* ATCC 25285, and *Eggerthella lenta* ATCC 43055. Plates for growth and aerotolerance controls were included.

*Multidrug resistance* (MDR) was defined as resistance to three or more families of antibiotics.

AZM and LEV were excluded from this analysis due to the lack of breakpoints for anaerobes. IMI and MOX were selected to represent carbapenems and fluoroquinolones, respectively.

### Statistical analysis

Data were analyzed using SPSS software, IBM SPSS Statistics version 21.0 (IBM Corp., Armonk, NY, USA).

Antimicrobial resistances were compared across the participating centers. For this evaluation, just one agent per

**Table 1** *In vitro* activity of 14 antimicrobial agents against 208 isolates of *Clostridioides difficile*.

Antimicrobial	Number of isolates	Range (µg/ml)	MIC50 (µg/ml)	MIC90 (µg/ml)	Geometric mean (µg/ml)	Breakpoint (S/I/R) (µg/ml)	% resistance	% intermed- iate	% suscepti- bility
Vancomycin	208	≤0.03–0.5	0.03	0.06	0.04	≤2/-/>>2 <sup>a</sup>	0*	-	100
Teicoplanin	207	≤0.03–2	0.03	0.03	0.03	≤2/-/>>2 <sup>c</sup>	0*	-	100
Metronidazole	207	≤0.12–2	0.5	1	0.38	≤8/16/≥32 <sup>b</sup>	0	0	100
Metronidazole						≤2/-/>>2 <sup>a</sup>	0*	-	100
Tigecycline	208	≤0.008–0.50	0.06	0.12	0.06	Not S >0.25 <sup>d</sup>	2.4*	-	97.6
Rifaximin	205	≤0.002 – >128	0.25	>128	0.56	≤32/-/>>32 <sup>e</sup>	44.4	-	55.6
Rifaximin						Not S >0.004 <sup>d</sup>	66.3*	-	33.7
Linezolid	208	≤0.03–8	1	4	1.03	≤4/-/>>4 <sup>c</sup>	4.8	-	95.2
Piperacillin/tazobactam	208	≤0.25–32	4	16	4.76	≤16/32–64/≥128 <sup>b</sup>	0	3.8	96.2
Meropenem	208	≤0.12–8	2	4	1.61	≤4/8/≥16 <sup>b</sup>	0	0.5	99.5
Imipenem	208	≤0.12–32	8	16	6.38	≤4/8/≥16 <sup>b</sup>	18.3	38.9	42.8
Ertapenem	207	≤0.12–32	4	16	4.24	≤4/8/≥16 <sup>b</sup>	10.6	28.5	60.9
Clindamycin	207	≤0.12 – >64	8	>64	11.51	≤2/4/≥8 <sup>b</sup>	55.6	14.5	29.9
Azithromycin	208	≤0.12 – >128	64	>128	30.39	≤2/-/>>2 <sup>c</sup>	82.2	-	17.8
Moxifloxacin	208	≤0.12–32	8	32	5.15	≤2/4/≥8 <sup>b</sup>	50.5	3.4	46.1
Moxifloxacin						Not S >4 <sup>d</sup>	50.5*	-	49.5
Levofloxacin	208	≤0.12 – >128	16	128	18.21	≤2/4/≥8 <sup>b</sup>	63.5	28.9	7.6
Levofloxacin						-/-/>>8 <sup>f</sup>	51.4*	-	48.6

Breakpoints were defined as susceptible, intermediately resistant or resistant with reference to CLSI, EUCAST or published data.

<sup>a</sup> EUCAST, clinical breakpoint Tables v. 13.0, anaerobic bacteria, *Clostridioides difficile* valid from 2023-01-01.

<sup>b</sup> CLSI, M100 Ed33, Table 2J – MIC breakpoint for anaerobes. 2023 (the breakpoint for moxifloxacin was used for levofloxacin).

<sup>c</sup> EUCAST, clinical breakpoint Tables v. 13.0, *Staphylococcus aureus* valid from 2023-01-01.

<sup>d</sup> EUCAST, clinical breakpoint Tables v. 9.0, *Clostridioides difficile* epidemiological cut-off values (ECOFF), 2019 (the ECOFF for rifampicin was used for rifaximin)<sup>9</sup>.

<sup>e</sup> Breakpoint from Reigadas et al.<sup>37</sup>.

<sup>f</sup> Breakpoint from Wang, Suo et al.<sup>49</sup>.

\* Non-susceptible, according to EUCAST epidemiological cut-off.

family was considered and strains from one of the centers (IL) were excluded because of the low number provided. The comparison was carried out using the chi-square test and statistical significance was set at  $p < 0.05$ .

## Results

Results of ranges of MIC, MIC<sub>50</sub>, MIC<sub>90</sub>, geometric means, and percentages of susceptibility, intermediate, and resistance for each antibiotic tested are shown in Table 1. The distributions of the isolates according to the MIC values of each agent are presented in Figure 1.

All isolates were susceptible to VAN and TEI (MIC  $\leq 2 \mu\text{g/ml}$ ). Similar results were observed for MTZ according to both CLSI and EUCAST breakpoints (MIC  $\leq 2 \mu\text{g/ml}$ ).

TIG was active against 97.6% of isolates, four isolates were non-susceptible with MICs of  $0.5 \mu\text{g/ml}$  for a breakpoint  $>0.25 \mu\text{g/ml}$ . One hundred and fourteen (114) isolates (55.6%) were susceptible to RFX according to the breakpoints defined by Reigadas et al.<sup>37</sup>; however, 91 of them (44.4%) were classified as non-susceptible when using the EUCAST ECOFF value<sup>9</sup>.

LNZ was active against 95.2% of the isolates tested, 10 isolates with MICs of  $8 \mu\text{g/ml}$  were classified as resistant for a breakpoint  $>4 \mu\text{g/ml}$ . Susceptibility to PTZ was observed in 96.2% of the isolates, and eight isolates, with MICs of  $32 \mu\text{g/ml}$ , were classified as intermediate.

MER was active against 99.5% of the isolates only one strain was classified as intermediate. Thirty-eight isolates were resistant to IMI, but none of them were resistant to ERT and MER.

Twenty-two isolates were resistant to ERT. Two of them were susceptible to IMI and 20 were intermediate. However, all 22 isolates were susceptible to MER. Seven isolates showed intermediate susceptibility to MOX according to CLSI guidelines, but were susceptible according to EUCAST breakpoints.

When comparing antibiotic resistance across centers, statistically significant differences ( $p < 0.05$ ) were found, mainly, for RFX, CLI, MOX, IMI, and AZM (Fig. 2).

RFX showed its characteristic bimodal behavior. Resistance ranged from 16.7% to 91.7% and was found mostly in two centers, HM and HP. The difference between these two institutions was 13.9% and was not significant; on the other hand, they were significantly different from the others.

For CLI, the range of resistance varied between 41.2% and 86.1%. The resistance rate of one center was significantly higher than the others.

MOX resistance ranged between 22.9% and 97.2%. HM and HP centers showed the highest values. Resistance in each of them differed significantly from the others.

Resistance to IMI varied from 9.4% to 55.6%. One center, with the highest resistance to IMI differed significantly from the others.

With regard to resistance to AZI, two groups presented statistically significant differences. In one group, CE and HG exhibited a resistance of 62.5% and 65.7%, respectively, whereas in the other group, HA, HC, HM, and HP showed values of 90.9%, 87.1%, 91.7%, and 97.2%, respectively.

MDR was detected in 80 isolates (38.5%), 63 (78.7%) were resistant to three families of antimicrobial agents,

and 17 (21.3%) were resistant to four. The most frequent combinations were RFX-MOX-CLI, which was present in 48 (60.0%) isolates, and RFX-IMI-MOX-CLI in 17 (21.3%) isolates. Other resistant combinations were RFX-MOX-IMI in 10 isolates (12.5%), IMI-MOX-CLI in 3 (3.8%) and RFX-IMI-CLI in 2 (2.5%). The distribution of MDR per center is shown in Figure 3.

## Discussion

CDI is a serious public health concern. *C. difficile* causes a variable spectrum of diseases, ranging from mild diarrhea to severe colon inflammation that can be life-threatening. Most patients develop diarrhea during or shortly after a course of antibiotic therapy.

Antibiotic resistance rates vary considerably in different studies, probably depending on geographic regions and local or national antibiotic policy<sup>43</sup>. Several studies documented the increase in the antibiotic resistance of *C. difficile* due to the inappropriate use of broad-spectrum agents, including cephalosporins, CLI, tetracycline, and fluoroquinolones, together with the emergence of hypervirulent strains<sup>13</sup> such as *C. difficile* BI/NAP1/027, and other ribotypes that were also associated with antibiotic resistance.

The antimicrobials recommended for CDI treatment are VAN, MET, or fidaxomicin<sup>25,46</sup>. It was shown that fidaxomicin treatment reduces the recurrence rate. Nevertheless, treatment failures and recurrences were reported. In countries like Argentina where fidaxomicin is not marketed, oral VAN and MET remain the agents of choice<sup>4</sup>.

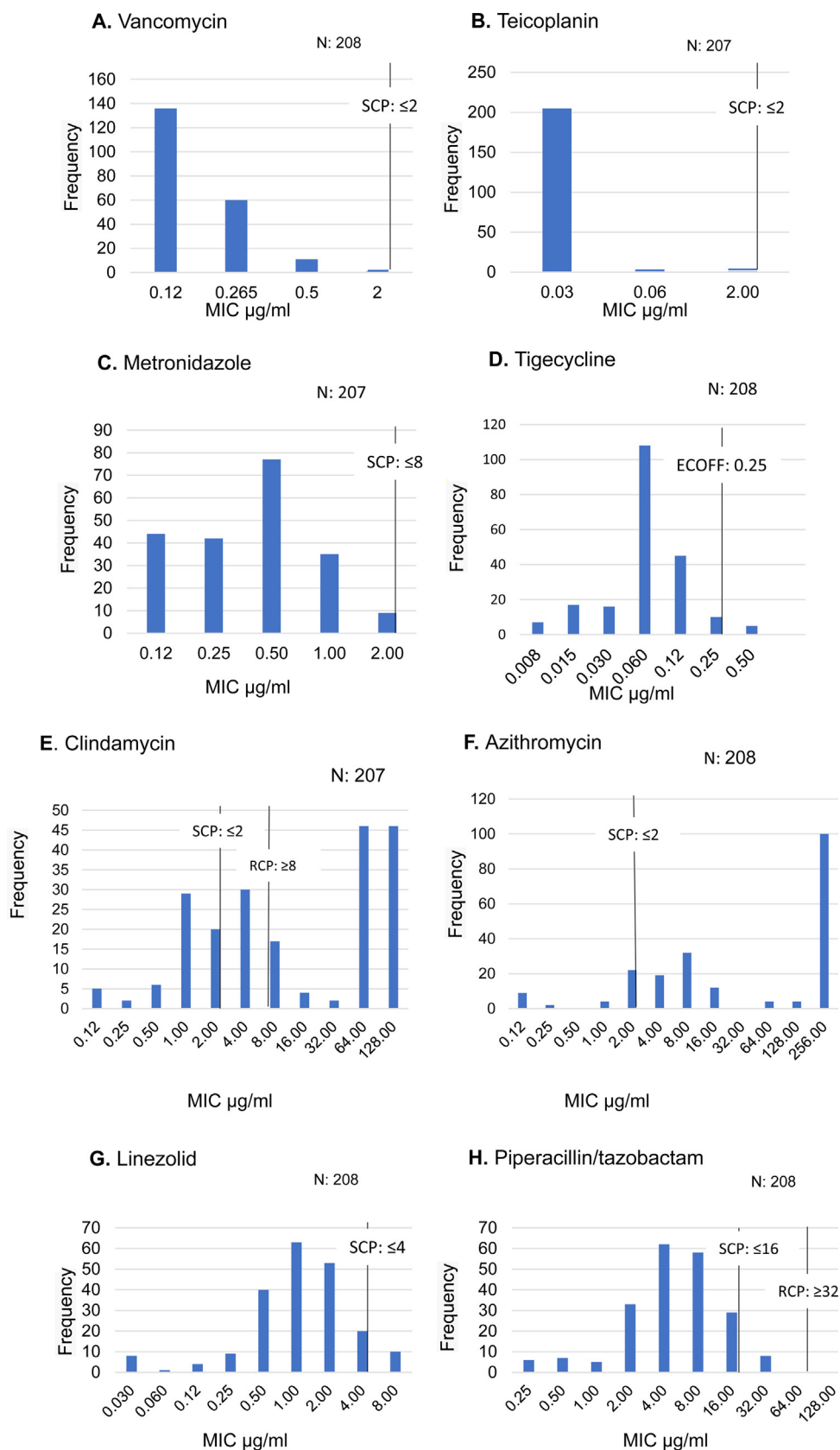
A systematic review of 39 studies found treatment failure in 22.4% and 14.2%, and a recurrence rate of 27.1% and 24.0% after treatment with MET and VAN, respectively<sup>47</sup>. Therefore, alternative treatments such as fidaxomicin, nitazoxanide, fusidic acid, TGC, TEI, and RXM were proposed<sup>6</sup>.

Appropriate antibiotic prescription is necessary to avoid the selection and spread of antibiotic-resistant *C. difficile* strains. Therefore, it is essential to conduct periodic antimicrobial susceptibility surveillance, with regional and institutional analyses to guide appropriate treatment in each area and institution. However, antimicrobial susceptibility testing (AST) is not routinely performed for *C. difficile*, and data evaluating MICs are limited, in particular from regions of Africa and South America<sup>13</sup>. In Argentina, there is still an insufficient number of reports regarding the susceptibility of circulating *C. difficile* strains. To date, and to the best of our knowledge, this is the largest survey published on the antimicrobial susceptibility of *C. difficile* conducted with isolates from human patients with CDI in Argentina.

## Vancomycin-teicoplanin

All our isolates were susceptible to VAN and TEI.

Although CLSI does not publish susceptibility breakpoints for VAN, EUCAST clinical breakpoints define an epidemiological cut-off value for non-susceptible *C. difficile* strains. Thus, according to EUCAST, all our isolates could be considered susceptible, which is consistent with unpublished local data from isolates obtained from pediatric<sup>32</sup> and adult patient<sup>28,38</sup>.



**Figure 1** Isolates distribution according to the MIC of each antimicrobial agent tested. SCP: susceptibility breakpoint; RCP: resistant breakpoint; ECOFF: epidemiological cut-off.

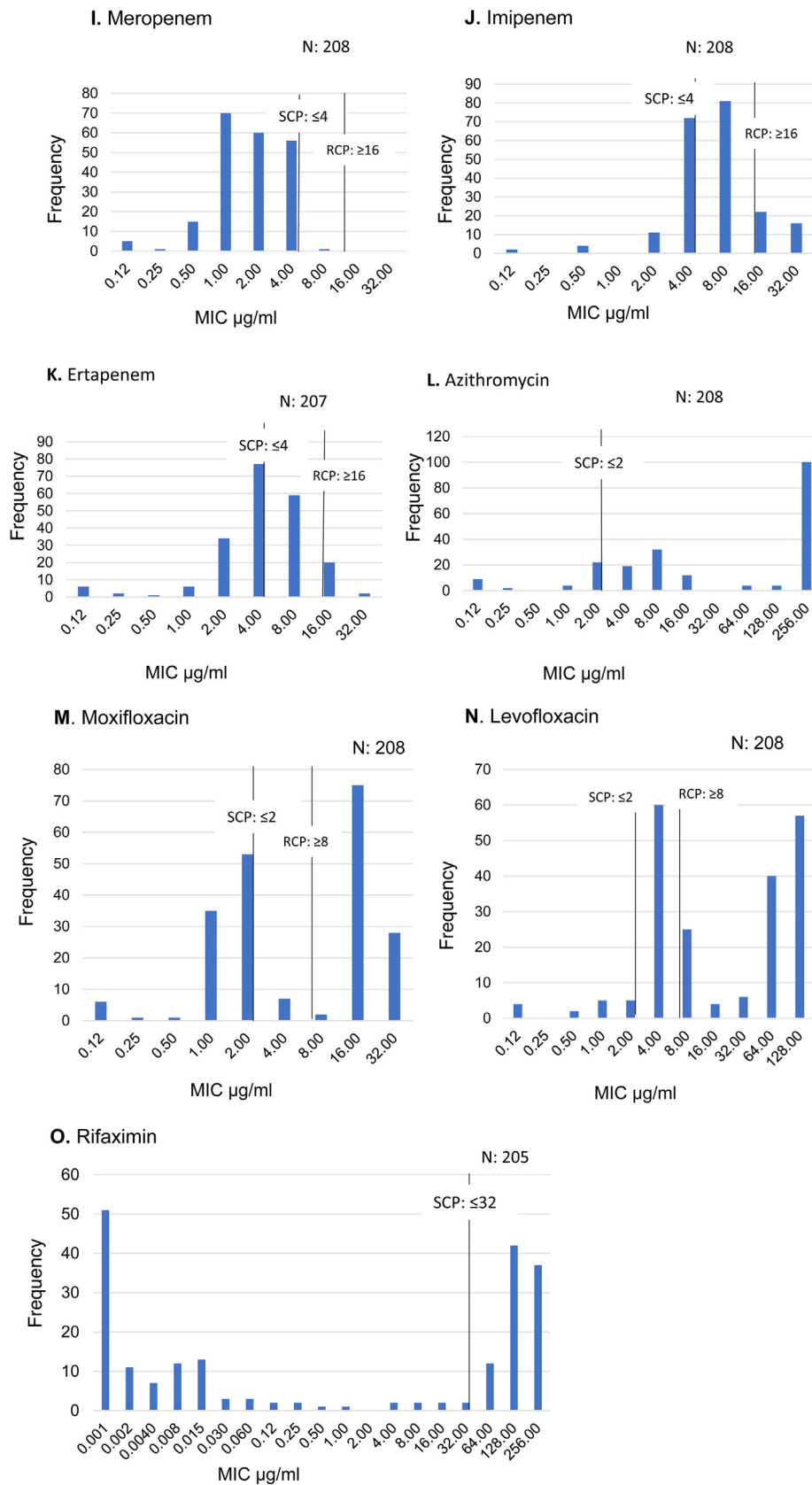
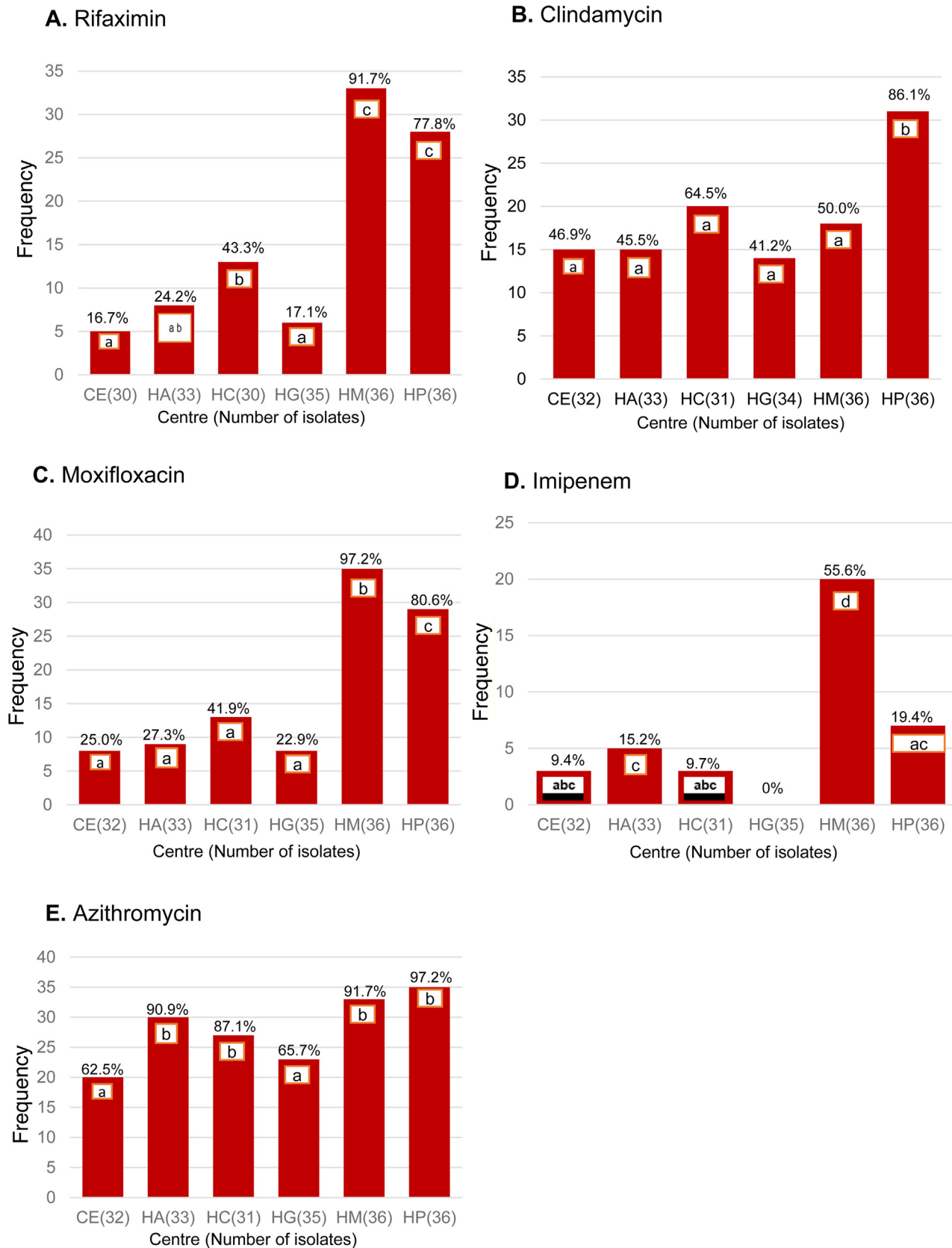
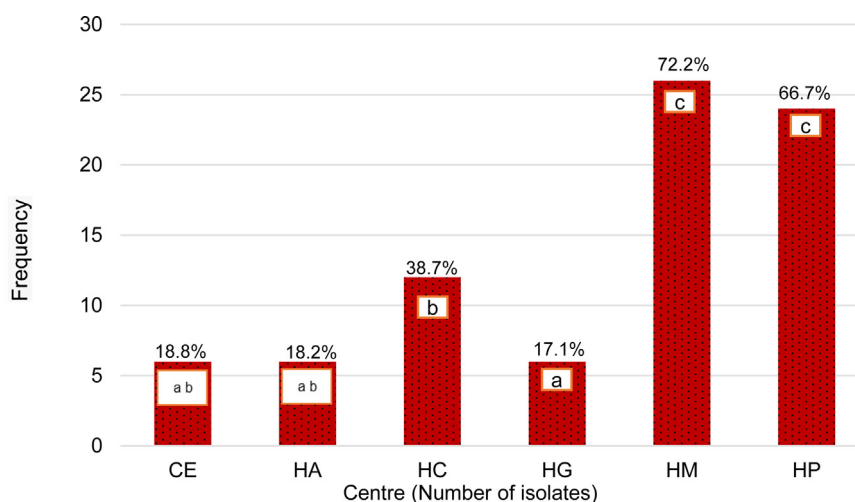


Figure 1 (Continued)



**Figure 2** Frequency of resistant isolates according to the centre of origin. Each letter over the resistance bars denotes a subset of centres whose proportions did not differ significantly from each other,  $p < 0.05$ .



**Figure 3** Distribution of multidrug resistance by centre. Each letter over the resistance bars denotes a subset of centres whose proportions did not differ significantly from each other,  $p < 0.05$ .

In 2016, while studying Brazilian isolates, Fraga et al.<sup>17</sup> found MIC<sub>50</sub> and MIC<sub>90</sub> of 4 µg/ml, with 58% of the isolates being non-susceptible. This important discrepancy compared with our results could be due to differences among the circulating strains, or to the use of the disk diffusion method for assessing antimicrobial susceptibility.

VAN was considered an agent with high and uniform *in vitro* activity, remaining stable over time<sup>14,20,41,43</sup>. However, a number of *C. difficile* strains with reduced susceptibility (MIC: >2, 4 and 8 µg/ml) were detected in isolates from Taiwan, Iran, China, and Israel<sup>43</sup>.

On the other hand, in a systematic review and meta-analysis, Saha et al.<sup>40</sup>, reported a significant VAN resistance increased by 3.6% after 2012. However, this finding could be affected by the significant heterogeneity in the susceptibility testing methodology. The highest VAN resistance rate was observed in the Americas, followed by Asia, and was significantly high among the RT027 strains. Other RTs displaying reduced VAN susceptibility were, RT017, RT078, RT126, RT001/072, and RT356<sup>18,34</sup>. The authors did not detect resistance to TEI.

In addition, plasmid-mediated resistance to VAN was described in a *C. difficile* isolate from a patient with CDI nonresponsive to VAN treatment<sup>27</sup>.

Clinical significance of reduced VAN susceptibility remains to be determined, since after oral treatment, VAN achieves high concentrations in feces (1000–3000 µg/ml), exceeding the highest MIC values recorded<sup>40,43</sup>.

Moreover, no VAN antimicrobial activity was detected on *C. difficile* spores, neither as a sporicidal nor as an inhibitor of sporulation, and the persistence of spores could contribute to the recurrence of CDI<sup>27</sup>. Furthermore, biofilm formation could be involved in VAN resistance<sup>34</sup>.

### Metronidazole

Resistance to MET remains rare but variable and dependent on the method and breakpoint considered<sup>11,17,40,43,44</sup>. It was reported to range from 0.11% to 18%<sup>11</sup>; however, resistant

strains have not yet been recorded in Argentina<sup>28,30,32,38</sup>. A review and meta-analysis from Sholeh et al.<sup>41</sup> described that when the EUCAST breakpoint was used, the highest MET-resistant rates were found in Asia (4.95%), followed by North America (3.95%) and Europe (1.95%). However, when the CLSI breakpoint was used, the highest resistance was observed in Europe (1.0%).

According to the CLSI and EUCAST breakpoints, 100% of our isolates were susceptible to this agent. Nine of them exhibited a MIC of 2 µg/ml, which coincides with the EUCAST epidemiological susceptibility cut-off value. In the same way, in Sao Paulo, Fraga et al.<sup>17</sup> did not find resistance among 50 isolates and reported MIC<sub>90</sub> of 2 µg/ml, one dilution higher than that found by us.

Gargis et al.<sup>21</sup> reported 97.3% susceptibility to MET in the USA when considering the EUCAST cut-off value but found no resistance when using the CLSI breakpoint. MIC<sub>50</sub> and MIC<sub>90</sub> values coincided with ours. Likewise, in a systematic review and meta-analysis from Mainland China in 2016, no MET-resistant strains were found<sup>44</sup>.

In 2022, Costa et al.<sup>11</sup> found that most isolates were susceptible to MET, but six isolates belonging to RT027 were resistant, whereas in 2018 Freeman et al.<sup>19</sup> reported an increase in susceptibility to MET, possibly due to a reduction in the use of this drug, or strain diversity<sup>11</sup>.

An Israeli paper<sup>45</sup> reported four (4.9%) isolates with reduced susceptibility, but using gradient strips methods and a breakpoint >2 µg/ml.

Additionally, in an Iranian study by Baghani et al.<sup>3</sup>, it was reported that more than half of the toxigenic *C. difficile* isolates exhibited reduced susceptibility to MET, with 30 isolates demonstrating high resistance with a MIC >256 µg/ml. The authors hypothesized that this high resistance could be linked to the overuse of MET. However, it is important to note that, unlike in our work, MICs were determined using test strips and interpreted according to EUCAST cut-off values.

MET resistance seems more common in non-toxigenic strains<sup>34</sup>. In Argentina, two isolates with an MIC of 16 µg/ml were detected<sup>48</sup>, and as previously reported, these strains were non-toxigenic.

Peláez et al.<sup>35</sup> documented that MET resistance is heterogeneous, inducible, and unstable after the freeze-thaw cycle. Although the true impact of heteroresistance is unknown, it could also influence treatment failure<sup>34,39</sup>.

Other factors such as iron concentration and resistant microbiota in the gut could also be related to MET resistance<sup>34</sup>. MET reaches low concentrations in the colon and feces, which may promote the development of resistance with impact on treatment failures<sup>34</sup>. In addition, as described for VAN, MET did not demonstrate significant inhibition of sporulation<sup>27</sup>. Furthermore, iron concentration, resistant microbiota in the gut, along with biofilm formation could influence the *in vivo* MET activity<sup>27,34</sup>, and explain cases of failure or recurrence associated to treatment with this drug.

### Tigecycline

A vast majority (97.6%) of the studied isolates was susceptible to TIG, but five (2.4%), coming from four different centers were classified as non-susceptible when applying the EUCAST epidemiological cut-off value<sup>16</sup>. A Brazilian study<sup>17</sup> reported values similar to ours, with MIC<sub>90</sub> of 0.12 µg/ml and MIC higher than 0.25 µg/ml in only one of 50 isolates. Likewise, in Europe a slightly reduced susceptibility to TGC was found; however, geometric mean MICs were more elevated in RT012<sup>20</sup>.

In an Israeli study, reduced susceptibility to TGC was found in 1.23% of isolates<sup>45</sup>; and the percentage of non-susceptible isolates was also low (1.6%) in a review and meta-analysis published in 2020<sup>41</sup>.

Supported by its *in vitro* activity, TGC was proposed as a potential alternative treatment of severe CDI. It inhibits spore formation and decreases toxin production, but it reduces the healthy microbiota of the gut and thus may contribute to CDI recurrence<sup>34</sup>. Furthermore, recently, the emergence of mobile TGC resistance genes has raised concern about the possible decrease in the activity of this agent and the need for controlled clinical studies to evaluate its efficacy and potential use, especially in patients for whom the treatments of choice have failed and fidaxomicin is not available<sup>34</sup>.

### Rifaximin

RFX is a rifamycin that has been used for intestinal decontamination and proposed as an adjunctive treatment to decrease recurrent CDI<sup>26,37</sup>. Our isolates showed a bimodal distribution (Fig. 1O), with more than 40% resistant ones and MIC<sub>90</sub> >128 µg/ml. Reigadas et al.<sup>37</sup> also found this type of distribution with a high value of CIM<sub>90</sub> >256 µg/ml, a geometric mean of 0.25 µg/ml, and 32.3% of resistance. Moreover, these authors demonstrated a good correlation between RFX results obtained by the agar dilution method and rifampicin results obtained by the E-test.

Lew et al.<sup>29</sup> tested RFX activity on *C. difficile* isolated from twelve Asia-Pacific countries. They identified a global resistance level of 15.5%, which is lower than that observed in this study. However, this resistance occurred mainly in RT017, which represented 67.7%. Although ribotyping was not performed in our study, Goorhies et al.<sup>23</sup> recognized the

circulation of RT017 in *C. difficile* isolates in a hospital in Argentina between 2000 and 2005. Other authors have also shown evidence of its circulation in recent years<sup>12,31</sup>.

Rifamycins in long-term regimes are commonly used in the treatment of tuberculosis, and previous exposure to rifampicin has been reported to be a risk factor for rifampicin-resistant *C. difficile*<sup>34,43</sup> strains. In the 2018 Freeman multicenter European study<sup>19</sup>, rifampicin resistance varied among the participating countries and was most notable in the Czech Republic (40.0–64.0%), Hungary (38.7–56.6%), Italy (36.6–47.0%), and Poland (5.0–44.0%). Later, the same authors<sup>20</sup> published annual changes between 13.5% and 11.8%, and increased resistance in RTs 027, 198, 018, 356, 017, and 176. Similarly, Lew et al.<sup>29</sup> found that RXM resistance varied across countries in the Asia-Pacific region and was absent in areas with low tuberculosis rates.

We also noted that the distribution of RXM resistance was heterogeneous across the participating centers. It varied between 16.7% and 91.7% and was highest in two of the seven centers (77.8% in HP and 91.7% in HM). In both, the number of patients with tuberculosis and, consequently, treated with rifampicin was greater than in the others. Rifampicin resistance in *C. difficile* isolates could be attributed to prior exposure to this agent with selection for resistant strains. However, transmission of rifampicin-resistant *C. difficile* strains to patients without a history of rifamycin use, could also occur<sup>34</sup>.

### Linezolid

Although *C. difficile* has been described as susceptible to LNZ, it has not been recommended for CDI treatment<sup>34</sup>. In this study, MIC varied between ≤0.03 and 8 µg/ml, and when using the EUCAST clinical breakpoint for *S. aureus*<sup>16</sup>, 10 isolates (4.81%) were resistant. This percentage falls within the range of resistance found in Europe, which spreads from 1% to 5.7%<sup>34</sup>.

### Piperacillin/tazobactam

In the current study, PTZ showed to be active against *C. difficile*. While 96% of the isolates were susceptible, none were resistant. Similarly, Saha et al. reported that *C. difficile* resistance was less than 1%<sup>40</sup>, and in 2022, a meta-analysis comprising 17 studies reported a WPR rate of 0%<sup>41</sup>.

### Carbapenems

MER was the most active carbapenem studied, and similarly to other publications<sup>7,41</sup> no resistant isolates were found. Then, ETP, which was active against 61% of the isolates, was 39% less active than MER, but more active than IMI. Information on the *in vitro* activity of carbapenems against *C. difficile* is scarce, and we did not find data on ETP activity; nonetheless, cases of ICD have been reported following treatment with this agent.

In our series, 18.27% of the isolates were resistant to IMI and only 42.79% were susceptible. However, the percentage of resistance was not uniform across the centers, finding a range of 46.20% (9.40–55.60%). European studies have

shown variable percentages of resistance to IMI between countries, with values ranging from 0% in Belgium, Bulgaria and Sweden, to 76% in Hungary<sup>19,24,36</sup>. Furthermore, an Iranian publication<sup>22</sup>, using the disk diffusion method, recorded resistance in 81% of *C. difficile* isolates. In Europe, IMI resistance was primarily associated with RT017<sup>24,36</sup>. The differences observed among our centers may also be attributed to ribotype dissemination; however, molecular studies were not included in our actual aims. Additional factors such as the use of carbapenems, differences in population demographics, and clonal dissemination within each institution could also contribute to these differences.

## Clindamycin

CLI administration is a high-risk factor for the development of CDI, as its resistance is widespread among *C. difficile* strains, varying between 25% and 96%<sup>33</sup>. In this survey, less than a third of the isolates were susceptible, and the global resistance was 55.6%.

High-level resistance to CLI has already been detected in Argentina<sup>28,38</sup>. In addition, the presence of the *ermB* gene was evidenced in local RT017 isolates<sup>23</sup>.

Our current resistant rate aligns with findings from a Pan-European survey<sup>18</sup> and two systematic reviews and meta-analysis<sup>13,41</sup>. The first reported a global resistance rate of 49.6%<sup>18</sup> while the WPR values from the meta-analysis were 59%<sup>41</sup> and 61%<sup>13</sup>, respectively.

On the other hand, our study found lower resistance rates to CLI compared with the 81.7% reported in a systematic review and meta-analysis from Mainland China<sup>44</sup>.

Sholeh et al.<sup>41</sup> identified significant regional differences, with a higher prevalence of resistance in Asia (72.9%) and South America (59%). In this study, when comparing data across different centers, one of them exhibited a CLI resistance rate of 86.1%, a value significantly higher than those observed in the other hospitals, but more in line with those reported in Mainland China.

## Azithromycin

Regarding AZM, 82% of our isolates exhibited resistance. While AZM is not used for treating CDI, the resistance observed may clarify its potential to induce the disease. Similarly, a study conducted on elderly individuals residing in a nursing home in Iran reported a high AZM resistance rate of 61%, when the disk diffusion method was used<sup>22</sup>.

## Moxifloxacin and levofloxacin

Fluoroquinolones are linked to an increased risk of CDI<sup>43,49</sup>, and resistance has been detected in 47% of *C. difficile* isolates<sup>43</sup>. Mutations in the GyrA and/or GyrB genes have been observed in these resistant strains<sup>21</sup>. Furthermore, MICs were found to be higher in isolates that contained the binary toxin<sup>11</sup>. However, in a study carried out in an Argentine pediatric hospital, resistance to fluoroquinolones was not associated to the binary toxin genes (personal communication from Dr. M. Litterio).

The rise in CDI associated with fluoroquinolones has coincided with an increasing incidence of *C. difficile* since the early 2000s<sup>19</sup>.

In this study, MOX was more active than LEV, and the difference was statistically significant ( $p=0.01$ ). However, despite the differences, both fluoroquinolones are potential risks for CDI. Resistance to MOX remained consistent regardless of the cut-off point used, whereas resistance to LEV was significantly higher when following the CLSI breakpoint. Moreover, LEV resistance was notably higher than the 20.3% reported by Wang et al.<sup>49</sup>.

The global MOX resistance found in this study was much higher than the 8% detected in Brazil in 2016<sup>17</sup>, and also surpassed the rates of 16%<sup>11</sup> and 22.9%<sup>21</sup> reported in the USA in 2022 and 2023, respectively. The resistance of our isolates was also higher than that of a Pan-European longitudinal surveillance, where MOX resistance was 39.9%. However, notable disparities were observed across countries, with a resistant range from 8% in France to 100% in Poland<sup>19</sup>. Moreover, a high resistance rate of 66% was found in Japan<sup>1</sup>.

The distribution of MOX resistance across our hospitals was heterogeneous; higher percentages were observed in two specific centers, HM and HP. These discrepancies were significant with respect to the other centers and also between each other. Figure 2 illustrates that the distribution of MOX-resistant strains mirrors that of RXM. These results suggest the presence of common factors in the institutions with the highest resistance levels. One potential factor could be the management of patients undergoing antituberculosis treatments, with the potential antibiotic selection pressure that this entails. Other factors may include clonal differences and the ribotypes present in these institutions.

## Multidrug resistance

MDR is commonly observed among *C. difficile* strains, and generally involves resistance to CLI, erythromycin, fluoroquinolones, cephalosporins, and tetracyclines<sup>34,42</sup>. Our isolates showed 38.5% of MDR to the tested agents; however, erythromycin, cephalosporins and tetracyclines were not evaluated. Thus, MDR could be underestimated in our series. The most frequent resistant combinations were RFX-MOX-CLI, and RFX-MOX-CLI-IMI. MDR prevalence varies by geographical location and strain type. In the USA, RT027 strains were particularly associated with MDR, and showed co-resistance to CLI, MOX, rifamycins, and tetracyclines<sup>34</sup>. European data indicate a predominance of MDR in RTs 017, 027, and 198, with percentages ranging from 28% to 77%<sup>34</sup>. In particular, Italian MDR strains were linked to RT356 and RT018, both characterized by resistance to CLI, erythromycin, MOX, and RIF<sup>43</sup>. While in the Asia-Pacific region, it was observed in 100% of RT369, 92.7% of RT018, and 66.1% of RT017<sup>34</sup>. In China, MDR varied from 5.8% in the Southwest to 43.75% in South China<sup>34</sup>.

As expected, we also confirmed variations among institutions. In line with RXM and MOX resistance, MDR was significantly higher in the centers with the highest tuberculosis prevalence.

In conclusion, this study demonstrated that VAN, TEI, and MET are the most active *in vitro* agents against *C. difficile* strains. Our results showed significant differences in

resistance for CLI, IMI, MOX, AZM, and RFX across centers. In particular, we highlight differences for RFX and MOX, which could be associated to the use of antituberculosis drugs.

We consider that this work has the strength of being a multicenter study, and of including a significant number of consecutive isolates from humans with CDI. This is the first publication and the most up-to-date information on the *in vitro* susceptibility of *C. difficile* in Argentina.

## Funding

The present work was supported by a grant awarded by IUCS-Fundación HA Barceló, Resolution No. 6973.

## Conflict of interests

The authors declare no conflicts of interest.

## Acknowledgement

We thank Dr. Ana Pereyra for providing necessary supply for the preparation of culture media.

## References

1. Aoki K, Takeda S, Miki T, Ishii Y, Tateda K. Antimicrobial susceptibility and molecular characterization using whole-genome sequencing of *Clostridioides difficile* collected in 82 hospitals in Japan between 2014 and 2016. *Antimicrob Agents Chemother.* 2019;63. <http://dx.doi.org/10.1128/AAC.01259-19>, e01259-19 [Epub 16 September 2019; PMID: 31527041; PMCID: PMC6879216].
2. Aptekorz M, Sacha K, Gofron Z, Kabala M, Harmanus C, Kuijper E, Martirosian G. Antibiotic resistance profile of RT 027/176 versus other *Clostridioides difficile* isolates in Silesia, Southern Poland. *Pathogens.* 2022;11:949. <http://dx.doi.org/10.3390/pathogens11080949>.
3. Baghani A, Ghourchian S, Aliramezani A, Yaseri M, Mesdaghinia A, Douraghi M. Highly antibiotic-resistant *Clostridium difficile* isolates from Iranian patients. *J Appl Microbiol.* 2018;125:1518–25.
4. Barcán L, Ducatenzeiler L, Bangher MC, Barcelona L, Cornistein W, Daciuk L, De Paula J, Desse J, Dictar M, Fernández-Canigia L, Nacinovich F, Scapellato P, Martínez JV, Jorge L, Obed M, Yantorno ML, Rollet R, Correa J, Maldonado ML, Predari SC, Legaria MC, Castello L, Rossetti MA, Vaustat D, Litterio M, Barberis C, Greco G, Abusamra L, Barletta J, Rodríguez Raimondo M, Brizuela M, Cuesta MCB, Ezcurra MC, Duarte A, Caeiro JP, Mena BS, Beltramo C, Sisto A, Smud A, Pereyra ML, Benito F, Bernachea MP, Farina J, Waldbaum C, Lambert S, Lescano ML, Soler Puy L, Valle S, Simioli F, Staneloni I, Maimone S, Alcalá W, Bejarano G, Baigorria MA, Nucetelli Y. Recomendaciones inter-sociedades para diagnóstico, tratamiento y prevención de las infecciones por *Clostridioides difficile*. *Medicina (Buenos Aires).* 2020;80:1–32. ISSN 1669-9106. [Accessed 8 Dec 2024]. Available from: [https://panel.aam.org.ar/img\\_up/03022020.1.pdf](https://panel.aam.org.ar/img_up/03022020.1.pdf)
5. Bishop EJ, Tiruvoipati R. Management of *Clostridioides difficile* infection in adults and challenges in clinical practice: review and comparison of current IDSA/SHEA, ESCMID and ASID guidelines. *J Antimicrob Chemother.* 2023;78:21–30.
6. Bouza E, Aguado JM, Alcalá L, Almirante B, Alonso-Fernández P, Borges M, Cobo J, Guardiola J, Horcajada JP, Masada E, Mensa J, Merchante N, Muñoz P, Pérez Sáenz JL, Pujol M, Reigadas E, Salavert M, Barberán J. Recommendations

- for the diagnosis and treatment of *Clostridioides difficile* infection: an official clinical practice guideline of the Spanish Society of Chemotherapy (SEQ), Spanish Society of Internal Medicine (SEMI) and the working group of Post-operative Infection of the Spanish Society of Anesthesia and Reanimation (SEDAR). *Rev Esp Quim.* 2020;33:151–75. <http://dx.doi.org/10.37201/req/2065.2020> [Epub 20 February 2020; PMID: 32080996; PMCID: PMC7111242].
7. Büchler AC, Rampini SK, Stelling S, Ledergerber B, Peter S, Schweiger A, Ruef C, Zbinden R, Speck RF. Antibiotic susceptibility of *Clostridium difficile* is similar worldwide over two decades despite widespread use of broad-spectrum antibiotics: an analysis done at the University Hospital of Zurich. *BMC Infect Dis.* 2014;14:607. <http://dx.doi.org/10.1186/s12879-014-0607-z> [PMID: 25425433; PMCID: PMC4247760.1].
8. Carroll KC, Bartlett JG. Biology of *Clostridium difficile*: implications for epidemiology and diagnosis. *Annu Rev Microbiol.* 2011;65:501–21.
9. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. Document M100. Wayne, EE.UU. 29th edition; 2019.
10. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. Document M100. Wayne, PA, EE.UU. 33rd edition; 2023.
11. Costa DVS, Pham NVS, Hays RA, Bolick DT, Goldbeck SM, Poulter MD, Hoang SC, Shin JH, Wu M, Warren CA. Influence of binary toxin gene detection and decreased susceptibility to antibiotics among *Clostridioides difficile* strains on disease severity: a single-center study. *Antimicrob Agents Chemother.* 2022;66. <http://dx.doi.org/10.1128/aac.00489-22>, e0048922 [Epub 21 July 2022; PMID: 35861541; PMCID: PMC9380565].
12. Crivaro AN, Carasi P, Salto I, Hugo A, Soldavini Peli-chotti PC, Bengoa A, Fragomeno M, Serradell MA, Minnaard J, Rolny I, Alul E, Arregui L, Fabra Martínez ME, Moreno Valero OJ, Facente A, Magariños F, Jewtuchowicz V, Pérez PF, Trejo FM. *Clostridioides difficile*: characterization of the circulating toxinotypes in an Argentinean public hospital. *Rev Argent Microbiol.* 2023;55:73–82. <http://dx.doi.org/10.1016/j.ram.2022.05.010>.
13. Dilnessa T, Getaneh A, Hailu W, Moges F, Gelaw B. Prevalence and antimicrobial resistance pattern of *Clostridium difficile* among hospitalized diarrheal patients: a systematic review and meta-analysis. *PLoS One.* 2022;17. <http://dx.doi.org/10.1371/journal.pone.0262597>, e0262597.
14. Eitel Z, Terhes G, Sóki J, Nagy E. Investigation of the MICs of fidaxomicin and other antibiotics against Hungarian *Clostridium difficile* isolates. *Anaerobe.* 2015;31:47–9.
15. European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Anaerobic bacteria. Version 13.0, valid from 2023-01-01.
16. European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Anaerobic bacteria. Version 9.0, valid from 2019-01-01.
17. Fraga EG, Nicodemo AC, Mello Sampaio JL. Antimicrobial susceptibility of Brazilian *Clostridium difficile* strains determined by agar dilution and disk diffusion. *Braz J Infect Dis.* 2016;20:476–81.
18. Freeman J, Vernon J, Morris K, Nicholson S, Todhunter S, Longshaw C, Wilcox MH, Pan-European Longitudinal Surveillance of Antibiotic Resistance Among Prevalent *Clostridium difficile* Ribotypes' Study Group. Pan-European longitudinal surveillance of antibiotic resistance among prevalent *Clostridium difficile* ribotypes. *Clin Microbiol Infect.* 2015;21. <http://dx.doi.org/10.1016/j.cmi.2014.09.017>, 248.e9–248.e16.
19. Freeman J, Vernon J, Pilling S, Morris K, Nicholson S, Shearman S, Longshaw C, Wilcox MH, Pan-European Longitudinal

- Surveillance of Antibiotic Resistance Among Prevalent *Clostridium difficile* Ribotypes Study Group. The ClosER study: results from a three-year Pan-European longitudinal surveillance of antibiotic resistance among prevalent *Clostridium difficile* ribotypes, 2011–2014. *Clin Microbiol Infect.* 2018;24:724–31.
20. Freeman J, Vernon J, Pilling S, Morris K, Nicolson S, Shearman S, Clark E, Palacios-Fabrega JA, Wilcox M, Pan-European Longitudinal Surveillance of Antibiotic Resistance among Prevalent *Clostridium difficile* Ribotypes' Study Group. Five-year Pan-European, longitudinal surveillance of *Clostridium difficile* ribotype prevalence and antimicrobial resistance: the extended ClosER study. *Eur J Clin Microbiol Infect Dis.* 2020;39:169–77, <http://dx.doi.org/10.1007/s10096-019-03708-7>.
21. Gargis AS, Karlsson M, Paulik AL, Anderson KF, Adamczyk M, Vlachos N, Kent AG, McAllister G, McKay SL, Halpin AL, Albrecht V, Campbell D, Korhonen LC, Elkins CA, Rasheed JK, Guh AY, McDonald LC, Lutgring JD, Emerging Infections Program Clostridioides difficile Infection Working Group. Reference susceptibility testing and genomic surveillance of *Clostridioides difficile* United States, 2012–7. *Clin Infect Dis.* 2023;76:890–6, <http://dx.doi.org/10.1093/cid/ciac817>.
22. Ghasemi A, Mohabati Mobarez A, Mostafavi E. Antibiotic susceptibility profile of *Clostridium difficile* bacteria isolated from older residents of a nursing home in Iran. *Iran J Ageing.* 2021;15:496–505, <http://dx.doi.org/10.32598/sija.15.4.2997.1> [Accessed 8 Dec 2024]. Available from: <http://salmandj.uswr.ac.ir/article-1-2052-en.html>
23. Goorhuis A, Legaria MC, van den Berg RJ, Harmanus C, Klaassen CH, Brazier JS, Lumelsky G, Kuijper EJ. Application of multiple-locus variable-number tandem-repeat analysis to determine clonal spread of toxin A-negative *Clostridium difficile* in a general hospital in Buenos Aires, Argentina. *Clin Microbiol Infect.* 2009;15:1080–6, <http://dx.doi.org/10.1111/j.1469-0691.2009.02759.x> [Epub 11 May 2009; PMID: 19438624].
24. Isidro J, Santos A, Nunes A, Borges V, Silva C, Vieira L, Mendes AL, Serrano M, Henriques AO, Gomes JP, Oleastro M. Imipenem resistance in *Clostridium difficile* ribotype O17, Portugal. *Emerg Infect Dis.* 2018;24:741–5, <http://dx.doi.org/10.3201/eid2404.170095> [PMID: 29553322; PMID: PMC5875251].
25. Johnson S, Lavergne V, Skinner AM, Gonzales-Luna AJ, Garey KW, Kelly CP, Wilcox MH. Clinical practice guideline by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA): 2021 focused update guidelines on management of *Clostridioides difficile* infection in adults. *Clin Infect Dis.* 2021;73, <http://dx.doi.org/10.1093/cid/ciab549>, e1029-e1044 [PMID: 34164674].
26. Johnson S, Schriever C, Galang M, Kelly CP, Gerding DN. Interruption of recurrent *Clostridium difficile*-associated diarrhea episodes by serial therapy with vancomycin and rifaximin. *Clin Infect Dis.* 2007;44:846–8.
27. Krutova M, Wilcox M, Kuijper E. *Clostridioides difficile* infection: are the three currently used antibiotic treatment options equal from pharmacological and microbiological points of view? *Int J Infect Dis.* 2022;124:118–23, <http://dx.doi.org/10.1016/j.ijid.2022.09.013>.
28. Legaria MC, Bianchini H, Castello L, Carloni C, di Martini A, Fernández Canigia L, Litterio M, Rollet R, Rossetti A, Predari SC. Primer consenso argentino para el estudio de la sensibilidad *in vitro* a los antimicrobianos de las bacterias anaerobias de importancia clínica en humanos. *Rev Argent Microbiol.* 2011;43:51–66, <http://dx.doi.org/10.1590/S0325-75412011000100012> [PMID: 21491069].
29. Lew T, Putsathit P, Sohn KM, Wu Y, Ouchi K, Ishii Y, Tateda K, Riley TV, Collins DA, Clostridium difficile Asia Pacific (CDAP) Study Group. Antimicrobial susceptibilities of *Clostridium difficile* isolates from 12 Asia-Pacific countries in 2014 and 2015. *Antimicrob Agents Chemother.* 2020;64, <http://dx.doi.org/10.1128/AAC.00296-20>, e00296-20.
30. Litterio MR, Roldán C, Angeloff M, D'Amore V, Hernández C. Sensibilidad antibiótica de *Clostridium difficile* aislados en un hospital pediátrico. Congreso internacional de infectología pediátrica y vacunas – SADIP; 2014 [Abstract ID24].
31. Lopardo B, Morfin-Otero R, Moran-Vazquez II, Noriega F, Zambrano G, Luxemburger C, Foglia G, Rivas EE. Epidemiology of *Clostridium difficile*: a hospital-based descriptive study in Argentina and Mexico. *Braz J Infect Dis.* 2015;19:8–14, <http://dx.doi.org/10.1016/j.bjid.2014.07.004> [Epub 29 August 2014; PMID: 25179510; PMID: PMC9425260].
32. Maldonado ML, Hernández C, Litterio M. *Clostridium difficile* susceptibility profiles to vancomycin and metronidazol in a pediatric population. *Int J Infect Dis.* 2018;73S:121, <http://dx.doi.org/10.1016/j.ijid.2018.04.3691> [Abstract UMP.023].
33. Nagy E. Anaerobic infections. Update on treatment considerations. *Drugs.* 2010;70:841–58.
34. O'Grady K, Knight DR, Riley TV. Antimicrobial resistance in *Clostridioides difficile*. *Eur J Clin Microbiol Infect Dis.* 2021;40:2459–78, <http://dx.doi.org/10.1007/s10096-021-04311-5>.
35. Peláez T, Cercenado E, Alcalá L, Marín M, Martín-López A, Martínez-Alarcón J, Catalán P, Sánchez-Somolinos M, Bouza E. Metronidazole resistance in *Clostridium difficile* is heterogeneous. *J Clin Microbiol.* 2008;46:3028–32.
36. Pituch H, Obuch-Woszczatyński P, Wultańska D, Nurzyńska G, Harmanus C, Banaszekiewicz A, Radzikowski A, Łuczak M, van Belkum A, Kuijper E. Characterization and antimicrobial susceptibility of *Clostridium difficile* strains isolated from adult patients with diarrhoea hospitalized in two university hospitals in Poland, 2004–2006. *J Med Microbiol.* 2011;60:1200–5, <http://dx.doi.org/10.1099/jmm.0.029801-0> [Epub 17 March 2011; PMID: 21415206].
37. Reigadas E, Muñoz-Pacheco P, Vázquez-Cuesta S, Alcalá L, Marín M, Martín A, Bouza E. Rifaximin-resistant *Clostridium difficile* strains isolated from symptomatic patients. *Anaerobe.* 2017;48:269–72.
38. Rollet R, Vaustat D, Cabrera R, Callejo R, Menéndez B, Hardie N. *Clostridium difficile*. Su sensibilidad *in vitro* frente a antibióticos de amplio uso en un hospital de enfermedades infecciosas. VII Congreso de la Sociedad Argentina de Bacteriología, Micología y Parasitología Clínicas. SADEBAC 2012, Abstract P127. *Rev Argent Microbiol.* 2012;44 Suppl. 1:56.
39. Rollet R, Vaustat D, Menéndez B, Hardie N. *Clostridium difficile*: su comportamiento *in vitro* frente a metronidazol. XIII Congreso Argentino de Microbiología. II Congreso Microbiología Agrícola y Ambiental 2013, Abstract O-005. *Rev Argent Microbiol.* 2013;45 Suppl. 1:7–8.
40. Saha S, Kapoor S, Tariq R, Schuetz AN, Tosh PK, Pardi DS, Khanna S. Increasing antibiotic resistance in *Clostridioides difficile*: a systematic review and meta-analysis. *Anaerobe.* 2019;58:35–46.
41. Sholeh M, Krutova M, Forouzes M, Mironov S, Sadeghifard N, Molaeipour L, Maleki A, Kouhsari E. Antimicrobial resistance in *Clostridioides (Clostridium) difficile* derived from humans: a systematic review and meta-analysis. *Antimicrob Resist Infect Control.* 2020;9:158, <http://dx.doi.org/10.1186/s13756-020-00815-5>.
42. Spigaglia P, Barbanti F, Mastrantonio P, on behalf of the European Study Group on Clostridium difficile (ESGCD), Ackermann G, Balmelli C, Barbut F, Bouza E, Brazier J, Delmée M, Drudy D, Kuijper E, Ladas H, Mastrantonio P, Nagy E, Pituch H, Poxton I, Rupnik M, Wullt M, Yücesoy M. Multidrug resistance in European *Clostridium difficile* clinical isolates. *J Antimicrob Chemother.* 2011;66:2227–34, <http://dx.doi.org/10.1093/jac/dkr292>.

43. Spigaglia P. Recent advances in the understanding of antibiotics resistance in *Clostridium difficile* infection. *Ther Adv Infect Dis*. 2016;3:23–42.
44. Tang C, Cui L, Xu Y, Xie L, Sun P, Liu C, Xia W, Liu G. The incidence and drug resistance of *Clostridium difficile* infection in Mainland China: a systematic review and meta-analysis. *Sci Rep*. 2016;6:37865, <http://dx.doi.org/10.1038/srep37865> [PMID: 27897206; PMCID: PMC5126672].
45. Tkhawkho L, Nitzan O, Pastukh N, Brodsky D, Jackson K, Peretz A. Antimicrobial susceptibility of *Clostridium difficile* isolates in Israel. *J Glob Antimicrob Resist*. 2017;10:161–4.
46. van Prehn J, Reigadas E, Vogelzang E, Bouza E, Hristea A, Guery B, Krutova M, Torbjorn Norén T, Allerberger F, Coia JE, Goorhuis A, van Rossen TM, Ooijevaar R, Burns K, Scharvik Olesen BR, Tschudin-Sutter S, Wilcox M, Vehreschild MJGT, Fitzpatrick F, Kuijper ED, The Guideline Committee of the European Study Group on Clostridioides difficile. European Society of Clinical Microbiology and Infectious Diseases: 2021 update on the treatment guidance document for *Clostridioides difficile* infection in adults. *Clinical Microbiol Infect*. 2021;27:S1–21.
47. Vardakas KZ, Polyzos KA, Patouni K, Rafailidis PI, Samonis G, Falagas ME. Treatment failure and recurrence of *Clostridium difficile* infection following treatment with vancomycin or metronidazole: a systematic review of the evidence. *Int J Antimicrob Agents*. 2012;40:1–8, <http://dx.doi.org/10.1016/j.ijantimicag.2012.01.004> [Epub 6 March 2012; PMID: 22398198].
48. Vaustat D, Cabrera R, Callejo R, Menéndez B, Hardie N, Rollet R. Sensibilidad a vancomicina y metronidazol de aislamientos de *Clostridium difficile* procedentes de pacientes asistidos en el hospital F.J. Muñiz. 3º Congreso de Enfermedades Endemoepidémicas. Hospital de Infecciosas ‘‘F.J. Muñiz’’. Año 14. *El Muñiz Hoy*. 2011; Suppl. 1:13. ISSN 032 9-7357.
49. Wang R, Suo L, Xia Chen H, Song LJ, Yun Shen Y, Ping Luo Y. Molecular epidemiology and antimicrobial susceptibility of *Clostridium difficile* isolated from the Chinese People’s Liberation Army General Hospital in China. *Int J Infect Dis*. 2018;67:86–91.